Total Syntheses of the AChE Inhibitors (±)-Arisugacins F and G

Sir:

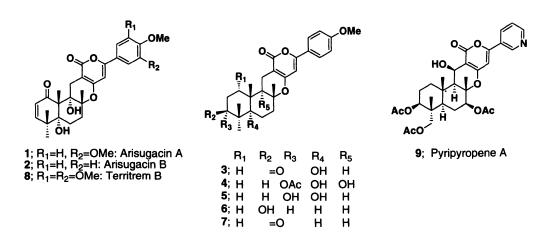
Arisugacins $A \sim G$ (1 \sim 7) were potent and selective inhibitors of acetylchlolinesterase (AChE) isolated by our group from a fermentation broth of Penicillium sp. FO- $4259^{1\sim5}$ together with the structurally related known compound, territrem B (8) (Fig. 1)^{6,7)}. Recently we disclosed the relative stereochemistries of them, and the absolute stereochemistries of 6 and $8^{8)}$, and the first total synthesis of (\pm) -arisugacin A (1), the most active congener^{9,10)}. Arisugacins F (6) and G (7), the simplest members of the family, are supposed to be the biosynthetic intermediates. Interestingly, structures $1 \sim 8$ resemble the pyripyropene A (9) which strongly inhibited acyl-CoA: cholesterol acyltransferase (ACAT), the enzyme that catalyzed intracellular esterification of cholesterol, and was isolated from Asperigillus fumigatus FO-1289 by $us^{11\sim 14}$. The first asymmetric total synthesis of pyripyropene A has also been accomplished via a convergent and efficient strategy¹⁵⁾. Herein, we describe the stereoselective and efficient total syntheses of arisugacins F (6) and G (7).

The ring system of the arisugacins suggests that a synthetic strategy through a biomimetic cyclization is possible. From the retrosynthetic perspective (Scheme 1), we envisioned that mercuric ion-induced cyclization of a polyolefin substrate (11) would provide the requisite the A, B, C-ring system of arisugacin skeleton (10). 10 would then be converted to 6 *via* pyrone annulation. Substrate 11 in turn could be derived from *trans*, *trans*-farnesyl bromide (12) and β -keto ester (13).

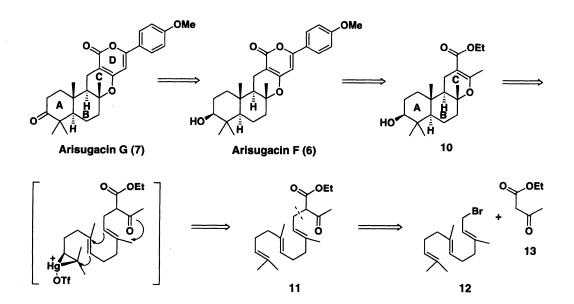
Ethyl acetoacetate (13) was alkylated with trans,

trans-farnesyl bromide (12) under a standard condition (NaH, DMF, rt) to give keto ester 11 in a 70% yield. Then, keto ester 11 was treated with mercury (II) trifluoromethanesulfonate ($Hg(OTf)_2$) and tetramethylurea (TMU) in CH₃CN, followed by sat. aq. NaCl to obtain the desired tricyclic organomercurial 14 as a single isomer 16,17 . This intermediate was converted to a mixture of the α - and β -hydroxylated stereoisomers 15 (32% yield) and 10 (18% yield) by means of the hydroxylation procedure (NaBH₄, O_2 , DMF)¹⁸⁾. The undesired α -hydroxylated isomer (15) was oxidized to ketone (TPAP, NMO, CH₂Cl₂), followed by the stereoselective reduction of the ketone to obtain the desired β -isomer 10. Next, dienolate γ -acylation and in situ cyclization^{19,20)} of **10** in the treatment with 3 eq. of LDA, and TMEDA, followed by addition of methyl pmethoxybenzoate (16) afforded the desired (\pm) -arisugacin F (6) in a 37% yield. Finally, the oxidation of 6 (TPAP, NMO, CH_2Cl_2) afforded the desired (±)-arisugacin G (7) in a 98% yield (Scheme 2). These synthetic arisugacins F and G were identical in all respects with natural 6 and 7 (400 MHz¹H- and 100 MHz¹³C-NMR, IR, HRMS, and TLC mobility in three solvent systems). Analytical data of 6: Rf=0.30 (Silica gel, Hexane: EtOAc=1:3), mp: 233~234°C (EtOAc), IR (KBr) $v \text{ cm}^{-1}$ 3449, 2937, 1690, 1637, 1571, 1513, 1405, 1257, 1182, 1123, ¹H-NMR (400 MHz, CDCl₃) δ : 7.72 (2H, d, J=9.0 Hz), 6.93 (2H, d, J=9.0 Hz), 6.25 (1H, s), 3.84 (3H, s), 3.24 (1H, dd, J=11.5, 4.5 Hz), 2.51 (1H, dd, J=17.0, 5.0 Hz), 2.22 (1H, dd, J=17.0, 13.0 Hz), 2.12 (1H, dt, J=12.0, 3.0 Hz), 1.81 (1H, dt, J=13.0, 3.5 Hz), 1.81 (1H, m), 1.71 (1H, m), 1.67 (1H, m), 1.63 (1H, m), 1.49 (1H, dd, J=13.0, 5.0 Hz), 1.43 (1H, m), 1.25 (3H, s), 1.11 (1H, dt, J=13.0, 4.0 Hz), 1.03 (3H, s), 0.99 (1H, dd, J=12.0, 2.0 Hz), 0.91 (3H, s),

Fig. 1. Structures of arisugacins $A \sim G(1 \sim 7)$, territrem B (8), and pyripyropene A (9).

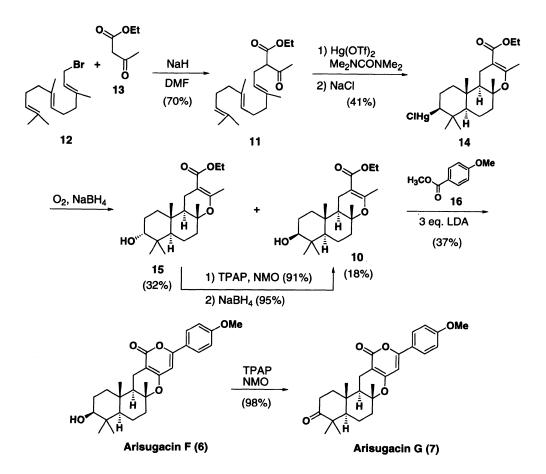


THE JOURNAL OF ANTIBIOTICS



Scheme 1. Retrosynthetic analysis of arisugacins F and G.

Scheme 2. Total syntheses of arisugacins F and G.



731

0.81 (3H, s), ¹³C-NMR (100 MHz, CDCl₃), δ : 164.7, 163.5, 161.5, 158.3, 127.0 (2C), 124.0, 114.2 (2C), 98.4, 96.7, 80.5, 78.5, 55.4, 55.0, 51.6, 40.4, 38.8, 37.4, 36.8, 28.1, 27.2, 20.7, 19.4, 17.2, 15.5, 15.1, HR-MS (EI) m/z: 438.2391 [M]⁺, Calcd for C₂₇H₃₄O₅: 438.2406 [M], Analytical data of 7; Rf=0.26 (Silica gel, Hexane: EtOAc=1:1), mp: 146~147°C (EtOAc), IR (KBr) $v \text{ cm}^{-1}$ 2943, 1703, 1638, 1574, 1513, 1403, 1257, 1181, 1120, ¹H-NMR (400 MHz, CDCl₃), δ: 7.73 (2H, d, J=9.0 Hz), 6.94 (2H, d, J=9.0 Hz), 6.26 (1H, s), 3.85 (3H, s), 2.60 (1H, ddd, J=16.0, 10.5, 7.5 Hz), 2.55 (1H, dd, J=17.0, 5.0 Hz), 2.46 (1H, ddd, J=16.0, 7.0, 3.5 Hz), 2.29 (1H, dd, J=17.0, 13.0 Hz), 2.16 (1H, m), 2.06 (1H, ddd, J=13.0, 7.5, 3.5 Hz), 1.74 (1H, m), 1.73 (1H, m), 1.58 (1H, m), 1.56 (1H, m), 1.55 (1H, m), 1.53 (1H, m), 1.30 (3H, s), 1.14 (3H, s), 1.07 (3H, s), 1.04 (3H, s), ¹³C-NMR $(100 \text{ MHz}, \text{ CDCl}_3) \delta$: 216.1, 164.5, 163.5, 161.5, 158.5, 127.0 (2C), 124.0, 114.2 (2C), 98.2, 96.6, 80.1, 55.4, 54.7, 51.0, 47.3, 39.8, 37.9, 36.7, 33.8, 26.6, 21.3, 20.6, 20.4, 17.3, 14.6, HR-MS (EI) m/z: 436.2251 [M]⁺, Calcd for C₂₇H₃₂O₅: 436.2250.[M].

In conclusion, the highly covergent, first total syntheses of the AChE inhibitors (\pm)-arisugacins F (6) and G (7) were completed by a very short sequence from the commercial available starting materials, ethyl acetoacetate (13), *trans*, *trans*-farnesyl bromide (12), and methyl *p*-methoxybenzoate (16), The key transformation, a polyolefin cyclization, provides rapid access to the vinylogous ester (14), establishing the A, B, C-ring system, which includes all five stereocenters of the target. Annulation of the α -pyrone ring in one step provided the natural product 6, and then oxidation afforded the natural product 7. The efficiency of this general approach suggests its exploitation in the synthesis of other members of the arisugacin class.

Acknowledgments

We are grateful to Ms. S. HIRANO for her helpful experiments and Mrs. N. SATO for measuring the NMR spectra. This study was supported in part by the Grant of the 21st Century COE Program, Ministry of Education, Culture, Sports, Science and Technology (MEXT), Scientific Research from the Ministry of Education, Science, Sports and Culture, Japan.

Masaki Handa Toshiaki Sunazuka Akihiro Sugawara Yoshihiro Harigaya Kazuhiko Otoguro Satoshi Ōmura*

Kitasato Institute for Life Sciences, Kitasato University, and The Kitasato Institute,

5-9-1 Shirokane, Minato-ku, Tokyo 108-8641, Japan

(Received April 17, 2003)

References

- ŌMURA, S.; F. KUNO, K. OTOGURO, T. SUNAZUKA, K. SHIOMI, R. MASUMA & Y. IWAI: Arisugacin, a novel and selective inhibitor of acetylcholinesterase from *Penicillium* sp. FO-4259. J. Antibiotics 48: 745~746, 1995
- KUNO, F.; K. OTOGURO, K. SHIOMI, Y. IWAI & S. ŌMURA: Arisugacins A and B, novel and selective acetylcholinesterase inhibitors from *Penicillium* sp. FO- 4259. I. Screening, taxonomy, fermentation, isolation and biological activity. J. Antibiotics 49: 742~747, 1996
- KUNO, F.; K. SHIOMI, K. OTOGURO, T. SUNAZUKA & S. ŌMURA: Arisugacins A and B, novel and selective acetylcholinesterase inhibitors from *Penicillium* sp. FO-4259. II. Structure elucidation. J. Antibiotics 49: 748~751, 1996
- 4) OTOGURO, K.; K. SHIOMI, Y. YAMAGUCHI, N. ARAI, T. SUNAZUKA, R. MASUMA, Y. IWAI & S. ŌMURA: Arisugacins C and D, novel acetylcholinesterase inhibitors and their related novel metabolites produced by *Penicillium* sp. FO-4259-11. J. Antibiotics 53: 50~57, 1996
- OTOGURO, K.; F. KUNO & S. OMURA: Arisugacins, selective acetylcholinesterase inhibitors of microbial origin. Pharmacol. Ther. 76: 45~54, 1997
- LING, K. H.; C.-K. YANG & F.-T. PENG: Territrems, tremorgenic mycotoxins of *Aspergillus terreus*. Appl. Environ. Microbiol. 37: 355~357, 1979
- LING, K. H.; H.-H. LIOU, C.-M. YANG & C.-K. YANG: Isolation, chemical structure, acute toxicity, and some physicochemical properties of territrem C from *Aspergillus terreus*. Appl. Environ. Microbiol. 47: 98~100, 1984
- HANDA, M.; T. SUNAZUKA, K. NAGAI, R. KIMURA, K. OTOGURO, Y. HARIGAYA & S. ŌMURA: Determination of absolute stereochemistries of arisugacin F and territrem B, novel acetylcholinesterase inhibitors. J. Antibiotics 54: 386~391, 2001
- 9) HANDA, M.; T. SUNAZUKA, K. NAGAI, R. KIMURA, T. SHIRAHATA, Z.-M. TIAN, K. OTOGURO, Y. HARIGAYA & S. ŌMURA: Convergent synthesis of arisugacin skeletons and their acetylcholinesterase inhibitory activity. J. Antibiotics 54: 382~385, 2001
- 10) SUNAZUKA, T.; M. HANDA, K. NAGAI, T. SHIRAHATA, Y. HARIGAYA, K. OTOGURO, I. KUWAJIMA & S. ŌMURA: The first total synthesis of (±)-arisugacin A, a potent, orally bioavailable inhibitor of acetylcholinesterase. Organic Letters 4: 367~369, 2002

- TOMODA, H.; Y. K. KIM, H. NISHIDA, R. MASUMA & S. ŌMURA: Pyripyropenes, novel inhibitors of acyl-CoA: cholesterol acyltransferase produced by *Aspergillus fumigatus*. I. Production, isolation, and biological properties. J. Antibiotics 47: 148~153, 1994
- 12) OMURA, S.; H. TOMODA, Y. K. KIM & H. NISHIDA: Pyripyropenes, highly potent inhibitors of acyl-CoA: cholesterol acyltransferase produced by *Aspergillus fumigatus*. J. Antibiotics 46: 1168~1169, 1993
- 13) KIM, Y. K.; H. TOMODA, H. NISHIDA, T. SUNAZUKA, R. OBATA & S. ÕMURA: Pyripyropenes, novel inhibitors of acyl-CoA : cholesterol acyltransferase produced by *Aspergillus fumigatus*. II. Structure elucidation of pyripyropenes A, B, C and D. J. Antibiotics 47: 154~162, 1994
- 14) TOMODA, H.; H. NISHIDA, Y. K. KIM, R. OBATA, T. SUNAZUKA, S. ŌMURA, J. BORDNER, M. GUADLLANA, P. G. DORMER & A. B. SMITH III: Relative and absolute stereochemistry of pyripyropene A, a potent, bioavailable inhibitor of acyl-CoA: cholesterol acyltransferase (ACAT). J. Am. Chem. Soc. 116: 12097~12098, 1994
- 15) NAGAMITSU, T.; T. SUNAZUKA, R. OBATA, H. TOMODA, H. TANAKA, Y. HARIGAYA & S. ŌMURA: Total synthesis of

(+)-pyripyropene A, a potent, orally bioavailable inhibitor of acyl-CoA : cholesterol acyltransferase. J. Org. Chem. $60: 8126 \sim 8127, 1995$

- 16) NISHIZAWA, M.; H. TAKENAKA & Y. HAYASHI: Experimental evidence of the stepwise mechanism of biomimetic olefin cyclization: trapping of cationic intermediates. J. Am. Chem. Soc. 107: 522~523, 1985
- 17) GOPALAN, A. S.; R. PRIETO, B. MULLER & D. PATERS: Polyene cyclizations using mercury (II) triflate-*N*,*N*dimethylaniline complex-participation by internal nucleophiles. Tetrahedron Lett. 33: 1679~1682, 1992
- 18) HILL, C. L. & G. M. WHITESIDES: Reactions of alkylmercuric halides with sodium borohydride in the presence of molecular oxygen. J. Am. Chem. Soc. 96: $870 \sim 876$, 1985
- PARKER, K. A. & L. RESNICK: The first total synthesis of a pyripyropene-type ACAT inhibitor, (+)-GERI-BP001.
 J. Org. Chem. 60: 5726~5728, 1995
- 20) OBATA, R.; T. SUNAZUKA, Z. TIAN, H. TOMODA, Y. HARIGAYA, S. ŌMURA & A. B. SMITH, III: New analogs of the pyripyropene family of ACAT inhibitors *via* α -pyrone fragmentation and γ -acylation/cyclization. Chemistry Letters 935~936, 1997